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Term:	L4 and urea		
Display:	<input type="text" value="10"/>	Documents in Display Format: <input type="text" value="-"/>	Starting with Number <input type="text" value="1"/>
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DB=USPT,EPAB,JPAB,DWPI; PLUR=YES; OP=ADJ

<u>L5</u>	L4 and urea	1	<u>L5</u>
<u>L4</u>	6492118.pn.	2	<u>L4</u>
<u>L3</u>	L2 and (urea near5 concentration\$1)	10	<u>L3</u>
<u>L2</u>	L1 and (hybridiz\$5 near5 temperature\$1)	113	<u>L2</u>
<u>L1</u>	hybridiz\$5 same buffer same urea	331	<u>L1</u>

END OF SEARCH HISTORY

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Term:

L3 and (low\$2 near5 temperature)

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DATE: Friday, August 13, 2004 [Printable Copy](#) [Create Case](#)**Set Name Query**

side by side

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result set

DB=USPT,EPAB,JPAB,DWPI; PLUR=YES; OP=ADJ

<u>L4</u>	L3 and (low\$2 near5 temperature)	19	<u>L4</u>
<u>L3</u>	L2 and array\$3	22	<u>L3</u>
<u>L2</u>	L1 and hybrid\$7	36	<u>L2</u>
<u>L1</u>	heat\$3 same probe same target same urea	36	<u>L1</u>

END OF SEARCH HISTORY

10001688

NSWER 1 OF 4 CAPLUS COPYRIGHT 2004 ACS on STN

AN 2003:757578 CAPLUS

DN 139:257687

TI Microcapillary hybridization chambers containing probes for detection of nucleic acids

IN Paszkowski, Jerzy; Guttman, Andras; Wang, Xun

PA Syngenta Participations Ag, Switz.

SO PCT Int. Appl., 30 pp.

CODEN: PIXXD2

DT Patent

LA English

FAN.CNT 1

	PATENT NO.	KIND	DATE	APPLICATION NO.	DATE
PI	WO 2003078045	A2	20030925	WO 2003-US7688	20030312
	WO 2003078045	A3	20040325		
	W:	AE, AG, AL, AM, AT, AU, AZ, BA, BB, BG, BR, BY, BZ, CA, CH, CN, CR, CU, CZ, DE, DK, DM, DZ, EE, ES, FI, GB, GD, GE, GH, GM, HR, HU, ID, IL, IN, IS, JP, KE, KG, KP, KR, KZ, LC, LK, LR, LS, LT, LU, LV, MA, MD, MG, MK, MN, MW, MX, MZ, NI, NO, NZ, PL, PT, RO, RU, SC, SD, SE, SG, SK, SL, TJ, TM, TR, TT, TZ, UA, UG, US, UZ, VC, VN, YU, ZA, ZW, AM, AZ, BY, KG, KZ, MD, RU, TJ, TM			
	RW:	GH, GM, KE, LS, MW, MZ, SD, SL, SZ, TZ, UG, ZM, ZW, AT, BE, BG, CH, CY, CZ, DE, DK, EE, ES, FI, FR, GB, GR, HU, IE, IT, LU, MC, NL, PT, RO, SE, SI, SK, TR, BF, BJ, CF, CG, CI, CM, GA, GN, GQ, GW, ML, MR, NE, SN, TD, TG			

PRAI US 2002-363869P P 20020312

AB The invention provides a microcapillary hybridization chamber made of a narrow bore tubing with probe segments. Each probe segment has oligonucleotide probes covalently attached to the inner wall of the tubing and the oligonucleotide probes within each segment have identical, known sequences. Many oligonucleotide probe segments can be present within a single centimeter of tubing. The invention further provides methods for using the microcapillary hybridization chambers in hybridization assays.

IT 50-00-0, Formaldehyde, biological studies 56-81-5, Glycerol, biological studies 57-13-6, Urea, biological studies 67-68-5, DMSO, biological studies 75-12-7, Formamide, biological studies 593-84-0, Guanidinium thiocyanate

RL: ARG (Analytical reagent use); BUU (Biological use, unclassified); ANST (Analytical study); BIOL (Biological study); USES (Uses)

(hybridization buffer comprising; microcapillary hybridization chambers containing probes for detection of nucleic acids)

L3 ANSWER 2 OF 4 MEDLINE on STN

DUPLICATE 1

AN 2002022961 MEDLINE

DN PubMed ID: 11464523

TI Fluorescence in situ hybridization method for co-localization of mRNA and GEP.

AU Oliva A A Jr; Swann J W

CS Oregon Health Sciences University, Portland, OR, Baylor College of Medicine, Houston, TX, USA.

NC HD24064 (NICHD)

NS18309 (NINDS)

NS34504 (NINDS)

NS37171 (NINDS)

SO BioTechniques, (2001 Jul) 31 (1) 74-6, 78-81.

Journal code: 8306785. ISSN: 0736-6205.

CY United States

DT Report; (TECHNICAL REPORT)

LA English

FS Priority Journals

EM 200112

ED Entered STN: 20020121

Last Updated on STN: 20020121

Entered Medline: 20011207

AB Co-localization studies using green fluorescent protein (GFP) and fluorescence immunohistochemistry have become commonplace. However, co-localization studies using GFP and mRNA in situ hybridization are rare, in large part because typical in situ hybridization reaction conditions often lead to the loss of GFP fluorescence. Here, we describe a new fluorescence mRNA in situ hybridization protocol using cRNA riboprobes that leaves GFP fluorescence intact. This protocol is based on a **urea-based hybridization buffer** and the Tyramide Signal Amplification system. This protocol should provide researchers engaged in the use of GFP with a solid starting point for adapting their own in situ hybridization protocols.

AB . . . fluorescence mRNA in situ hybridization protocol using cRNA riboprobes that leaves GFP fluorescence intact. This protocol is based on a **urea-based hybridization buffer** and the Tyramide Signal Amplification system. This protocol should provide researchers engaged in the use of GFP with a solid. . .

L3 ANSWER 3 OF 4 EMBASE COPYRIGHT 2004 ELSEVIER INC. ALL RIGHTS RESERVED.
on STN

AN 80152269 EMBASE

DN 1980152269

TI Gene mapping of cytoplasmic polyhedrosis virus of silkworm by the full-length mRNA prepared under optimized conditions of transcription in vitro.

AU Smith R.E.; Furuichi Y.

CS Roche Inst. Molec. Biol., Nutley, N.J. 07110, United States

SO Virology, (1980) 103/2 (279-290).

CODEN: VIRLAX

CY United States

DT Journal

FS 047 Virology

022 Human Genetics

LA English

AB Viral mRNA synthesis by the RNA polymerase associated with purified cytoplasmic polyhedrosis virus (CPV) was studied. The formation of full-length mRNA products was facilitated by including in the reaction mixture 100 mM sodium acetate, high concentrations of ribonucleoside triphosphates, and proteinase K. The 10 different species of CPV mRNAs were resolved into 9 discrete RNA bands by agarose gel electrophoresis at pH 3.5 in **buffer** containing 7 M **urea**. Each purified viral mRNA **hybridized** specifically to one of the viral genome segments which were separated by polyacrylamide gel electrophoresis into the 10 species of dsRNA. The relationship between the genome segments and their cognate mRNAs synthesized in vitro is thus established. Under optimal conditions of mRNA synthesis each of the genome segments was transcribed at a similar rate as determined from the yield of individual separated mRNA species. A recycling model of genome-associated RNA polymerase for viral transcription is discussed.

AB . . . different species of CPV mRNAs were resolved into 9 discrete RNA bands by agarose gel electrophoresis at pH 3.5 in **buffer** containing 7 M **urea**. Each purified viral mRNA **hybridized** specifically to one of the viral genome segments which were separated by polyacrylamide gel electrophoresis into the 10 species of. . .

L3 ANSWER 4 OF 4 CAPLUS COPYRIGHT 2004 ACS on STN

AN 1967:497251 CAPLUS

DN 67:97251

TI Urea-mediated freeze-thaw hybridization of lactate dehydrogenase

AU Massaro, Edward J.

CS Yale Univ., New Haven, CT, USA

SO Biochimica et Biophysica Acta (1967), 147(1), 45-51

CODEN: BBACAQ; ISSN: 0006-3002

DT Journal

LA English

AB The subunits of lactate dehydrogenases from a wide variety of related and unrelated organisms can be reassocd. in vitro into functional "hybrid molecules." This can be accomplished by subjecting isozyme mixts. to a freeze-thaw cycle in phosphate buffer containing the necessary hybridization-promoting substances: I-, Br-, Cl-, thiocyanate, or 1-anilinonaphthalene-8-sulfonate. It has been observed now that low concns. of **urea** (<0.1M) in Na phosphate **buffer** also will promote freeze-thaw **hybridization**. Urea and the other hybridization-promoting substances appear to function by a similar mechanism. It has been suggested that urea disrupts protein structure by an ion-exchange mechanism. It has also been observed that Tris not only inhibits freeze-thaw hybridization, but also protects the enzyme against freeze-thaw destruction. Under various conditions, the isozymes possessing the least net neg. charge are more susceptible to irreversible freeze-thaw denaturation than the other isozymic types. This may be the result of evolutionarily preserved structural characteristics common to all isozymic forms.

AB The subunits of lactate dehydrogenases from a wide variety of related and unrelated organisms can be reassocd. in vitro into functional "hybrid molecules." This can be accomplished by subjecting isozyme mixts. to a freeze-thaw cycle in phosphate buffer containing the necessary hybridization-promoting substances: I-, Br-, Cl-, thiocyanate, or 1-anilinonaphthalene-8-sulfonate. It has been observed now that low concns. of **urea** (<0.1M) in Na phosphate **buffer** also will promote freeze-thaw **hybridization**. Urea and the other hybridization-promoting substances appear to function by a similar mechanism. It has been suggested that urea disrupts protein structure by an ion-exchange mechanism. It has also been observed that Tris not only inhibits freeze-thaw hybridization, but also protects the enzyme against freeze-thaw destruction. Under various conditions, the isozymes possessing the least net neg. charge are more susceptible to irreversible freeze-thaw denaturation than the other isozymic types. This may be the result of evolutionarily preserved structural characteristics common to all isozymic forms.

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